

Pictures by: Ruchika Kashyap

# SENSITIVITY OF *PHOMOPSIS* SPECIES TO FLUXAPYROXAD, PYRACLOSTROBIN, AND TEBUCONAZOLE FUNGICIDES

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# OUTLINE



Justification for  
research



Methodology



Results and  
Conclusion



References



Acknowledgement



# JUSTIFICATION



## FRAC Group 3

DMI - Sterol biosynthesis inhibitor  
(SBI) fungicides

## FRAC Group 7

SDHI - Inhibitor of respiration in  
complex II at SDH

## FRAC Group 11

QoI - Inhibitor of respiration in  
complex III at Qo-site

# JUSTIFICATION



Classified under medium to high risk of resistance development (FRAC 2021)



For effective use of fungicides, it is crucial to monitor the fungicide sensitivity of fungal populations before chemical failures

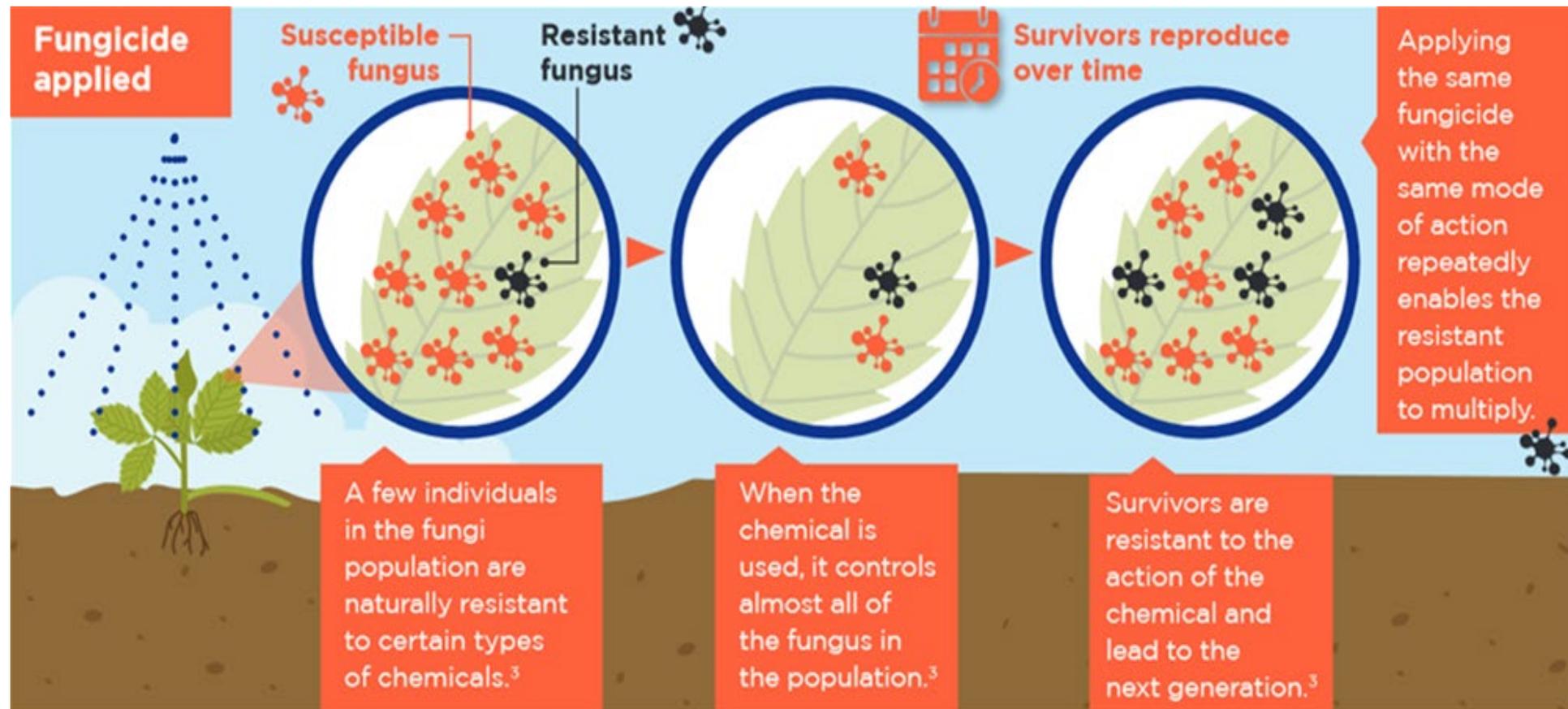


Monitoring will ensure prolonged and proper use of fungicides

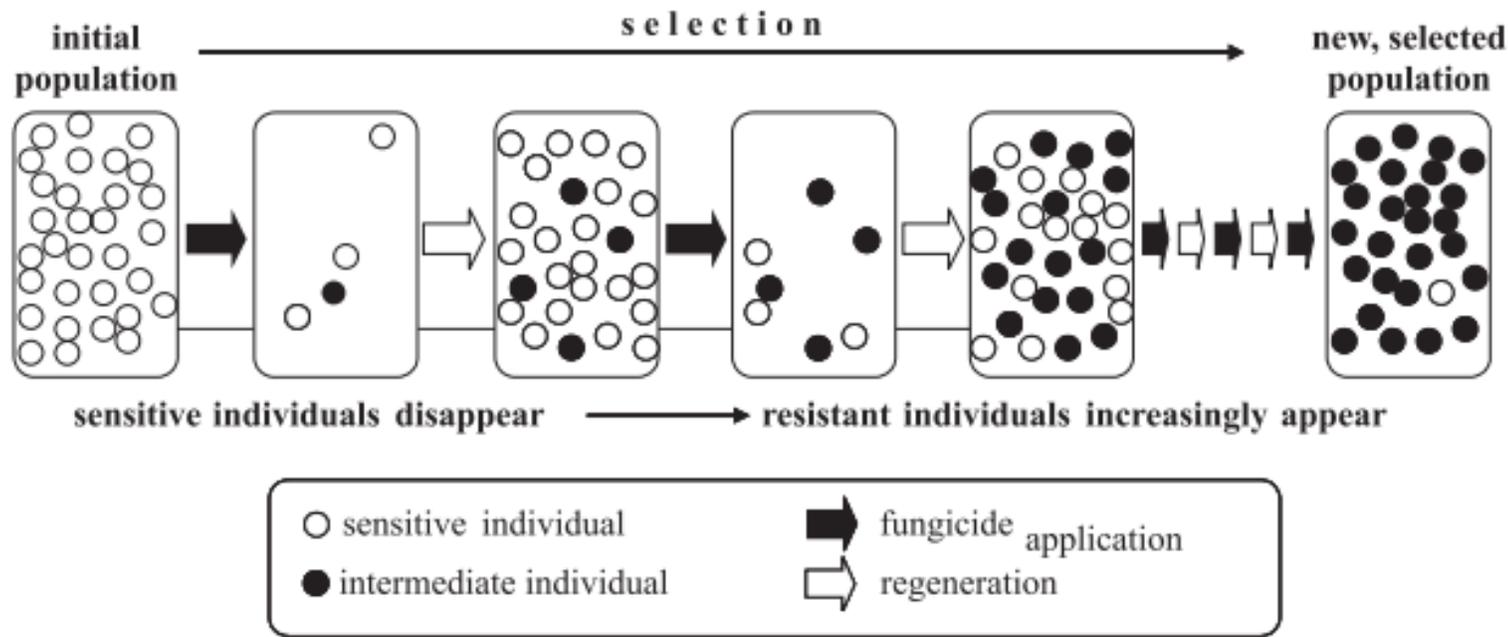


# WHAT IS FUNGICIDE RESISTANCE?

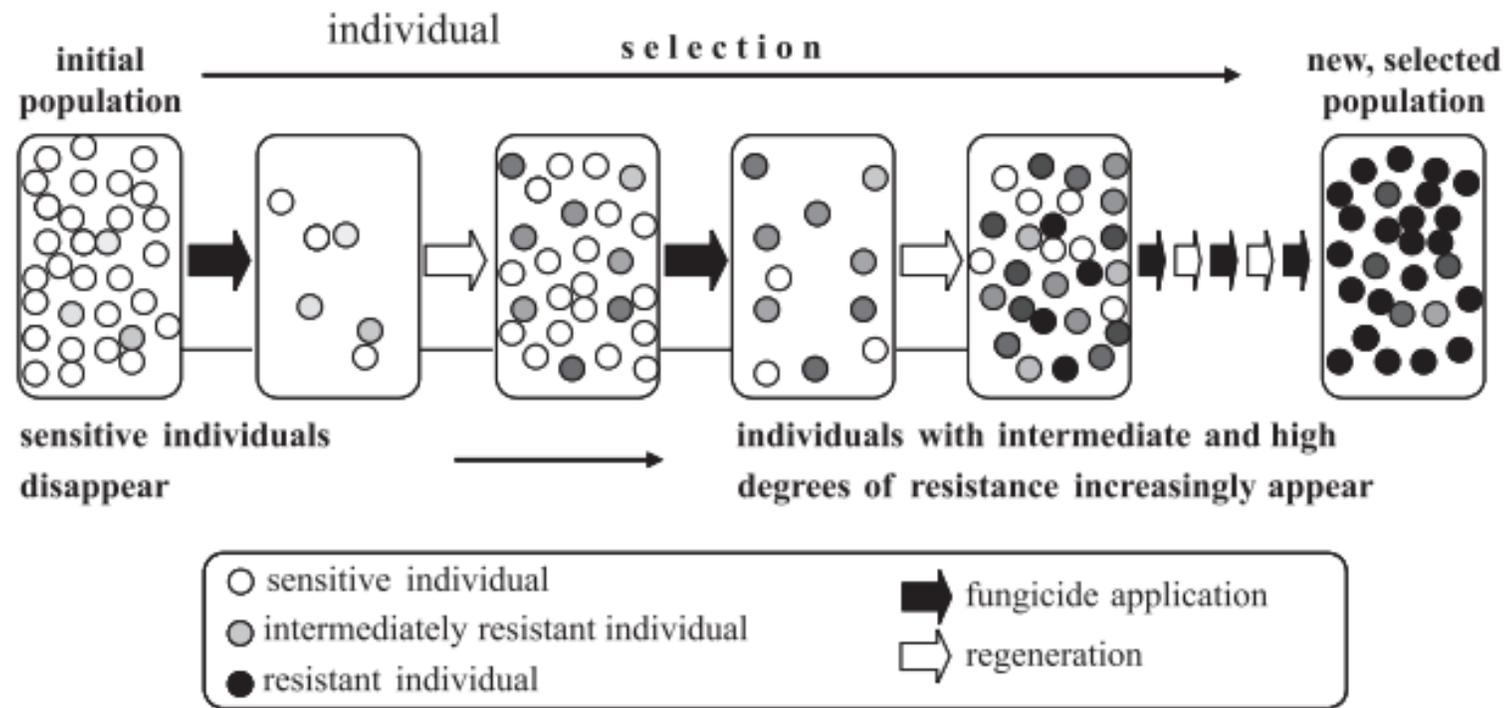
Refers to an acquired, heritable reduction in sensitivity of a fungus to a specific anti-fungal agent (or fungicide) (FRAC 2021)



A



B



# Types of Resistance

- Qualitative**
- Mutation-based
  - Discrete/Disruptive

- Quantitative**
- Gradual/Multistep
  - Continuous

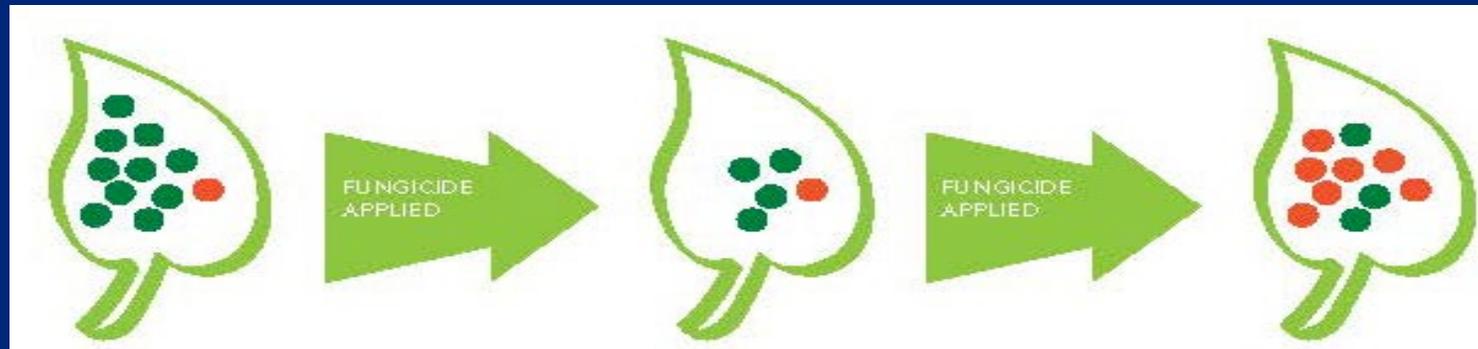
**Emergence of resistant population based on the type of resistance**

**(Deising et al. 2008)**



# RESEARCH OBJECTIVE

Determine the sensitivity of *D. gulyae* and *D. helianthi* to fluxapyroxad (FRAC 7), pyraclostrobin (FRAC 11), tebuconazole (FRAC 3), and fungicides *in vitro*

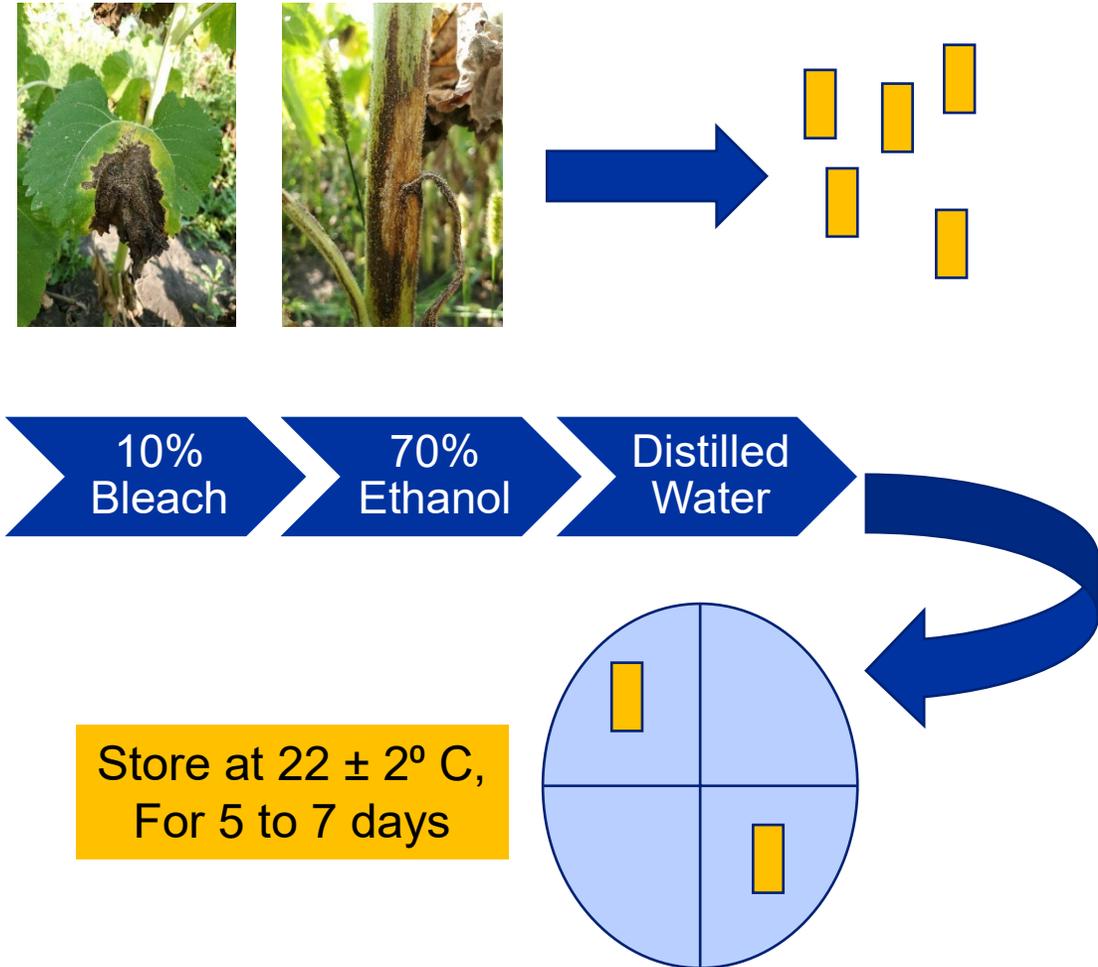


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# METHODOLOGY

## Isolate Collection

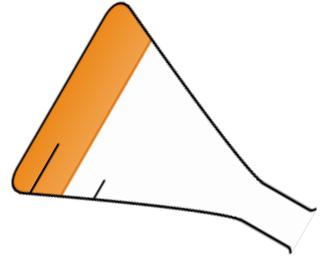
- **Number of isolates:** 52 isolates of *D. gulyae* and 54 isolates of *D. helianthi*
- **Locations:** Minnesota ( $n=31$ ), Nebraska ( $n=6$ ), North Dakota ( $n=30$ ), South Dakota ( $n=33$ ), Unknown ( $n=3$ )
- **Years:** 2013 to 2020
- **Baseline isolates:** One - *D. gulyae*, ex-type BRIP 54025 (Australia) and two – *D. helianthi*, 201540 (Former Yugoslavia) and 52763 (Texas)



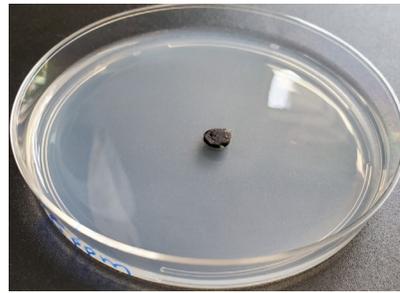
# METHODOLOGY

- Water agar serially amended with fungicides at different concentrations
- Colony diameter of each isolate measured twice after 5 days of incubation in dark at  $22\pm 2^{\circ}\text{C}$ .
- Experiment arranged in a completely randomized design with four plates (replications) for each fungicide concentration.

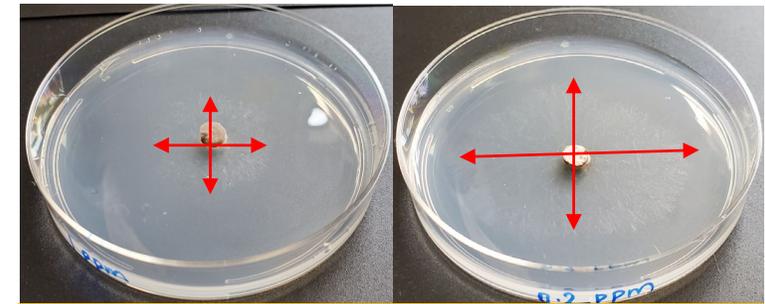




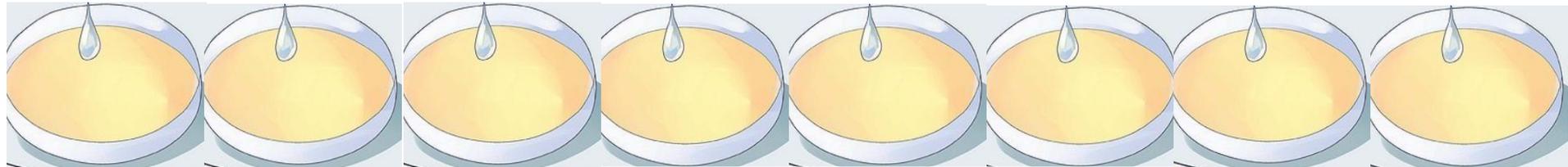
Fungicide amended media



6 mm fungal plug transferred to each concentration



Mycelial growth of fungus measured at right angles using scale at 5 to 7 days



Fluxapyroxad ( $\mu\text{g a.i./ml}$ )	0	0.001	0.01	0.1	1	10	100	-
Pyraclostrobin + SHAM ( $20 \mu\text{g a.i./ml}$ )	0	0.001	0.01	0.1	1	10	-	-
Tebuconazole ( $\mu\text{g a.i./ml}$ )	0	0.01	0.02	0.04	0.2	1	5	20

# ADDITION OF SALICYLHYDROXAMIC ACID (SHAM)

- When the normal respiration pathway is inhibited by Qols *in vitro*, fungus activates an alternate mitochondrial respiration pathway (Kaneko and Ishii 2009)
- SHAM inhibits the alternate pathway



# EFFECT OF SHAM ON MYCELIAL GROWTH OF *PHOMOPSIS SPECIES*

- Five isolates of each of *D. gulyae* and *D. helianthi* evaluated by dissolving SHAM in 0.1% (v/v) methanol for 50, 100, and 150 µg/ml
- Control plates included water agar amended with methanol and without methanol
- Analysis of variance compared SHAM concentrations, fungal isolates, and their interactions in R (R Core Team 2013)

Our preliminary study showed that the concentration, isolate, and the interaction effect was significant ( $p < 0.05$ )



# EFFECT OF SHAM ON MYCELIAL GROWTH OF *PHOMOPSIS* SPECIES IN ADDITION OF PYRACLOSTROBIN

- Ten isolates each of *D. gulyae* and *D. helianthi* evaluated to determine whether SHAM affected fungal growth in the presence of pyraclostrobin
- SHAM effect was evaluated at 20 and 100 µg/ml in combination with final concentrations of pyraclostrobin of 0.001, 0.01, 0.1, 1, 10 µg a.i./ml
- Plates amended with only SHAM served as control



# INHIBITORY EFFECT OF SHAM

Control plates with SHAM at 100  $\mu\text{g/ml}$  substantially inhibited the mycelial growth so SHAM at 20  $\mu\text{g/ml}$  was used for further studies

- Shi et al. (2020) – found significant growth inhibition of *Phomopsis asparagi* at SHAM  $\geq 40 \mu\text{g/ml}$
- Liang et al. (2015) found an apparent toxic effects of SHAM  $\geq 20 \mu\text{g/ml}$  on mycelial growth of *Sclerotinia sclerotiorum*.



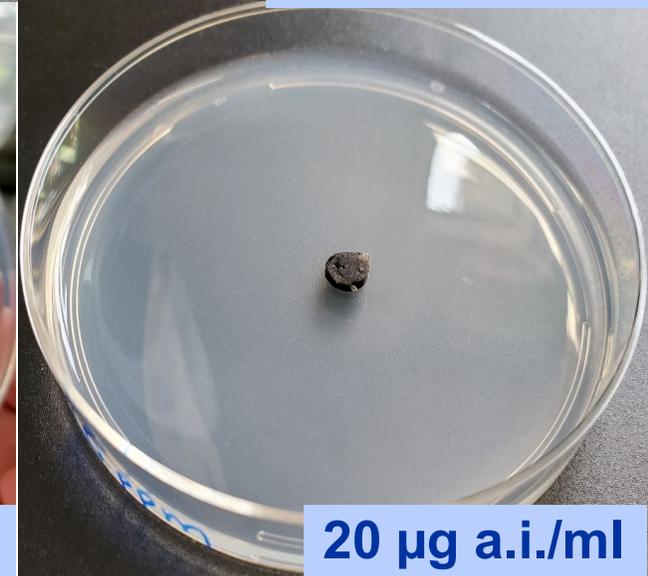
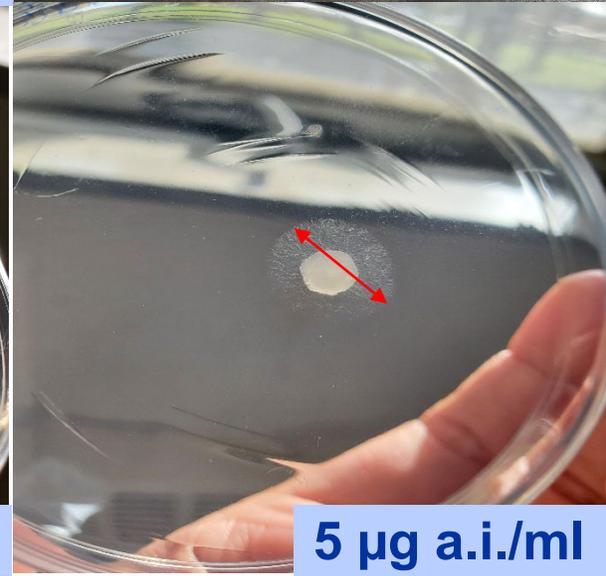
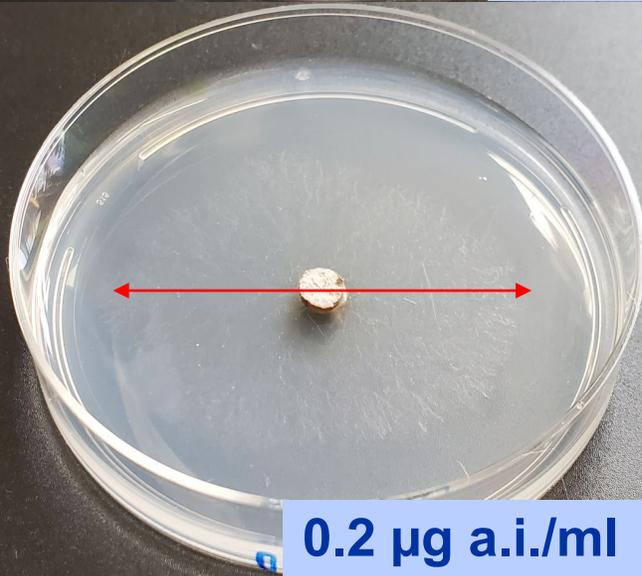
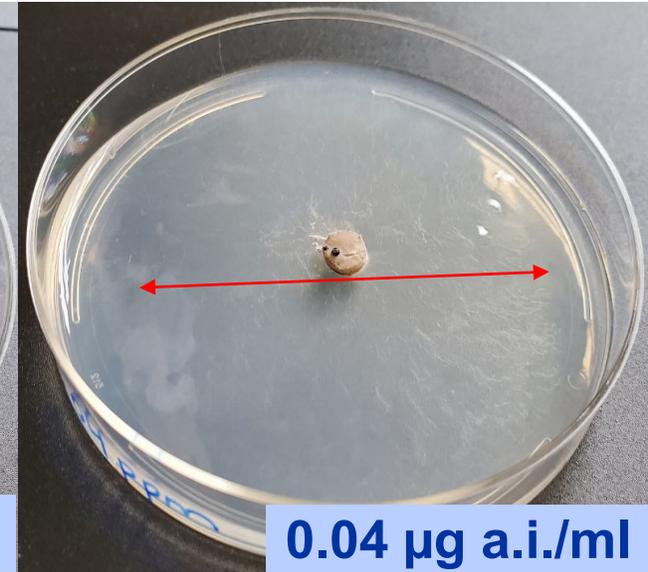
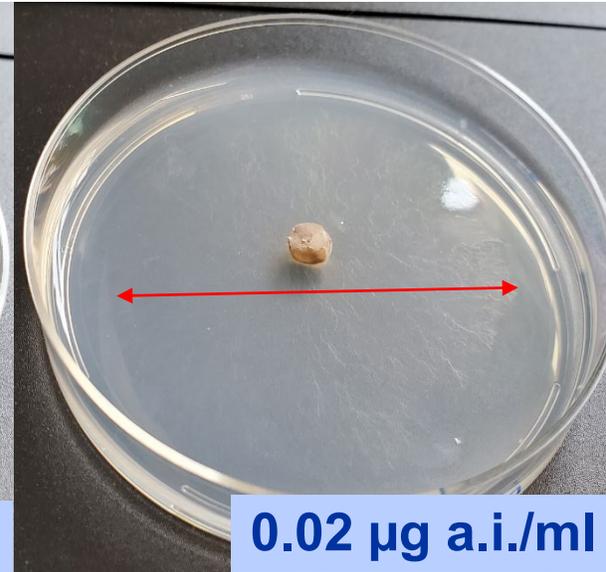
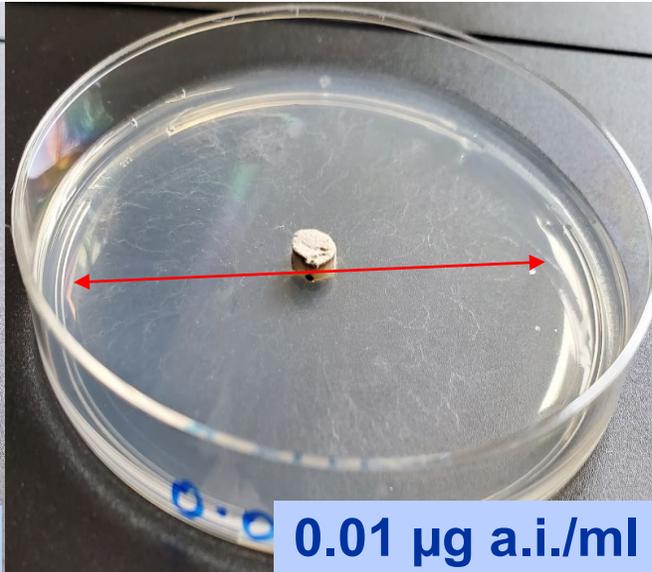
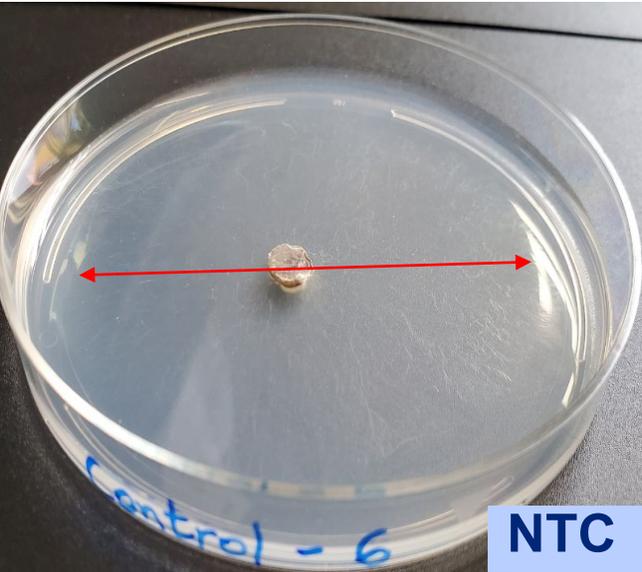
# T-TEST FOR PYRACLOSTROBIN AMENDED WITH AND WITHOUT SHAM

Species	EC50 ( $\mu\text{l/ml}$ )		T-value	p-value
	Without SHAM	With SHAM (20 $\mu\text{l/ml}$ )		
<i>D. gulyae</i>	0.639	2.097	0.844	0.410
<i>D. helianthi</i>	1.566	0.019	-2.990	0.004

Pyraclostrobin + SHAM (20 $\mu\text{g a.i./ml}$ )	0	0.001	0.01	0.1	1	10	-	-
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# SENSITIVITY OF *Diaporthe gulyae* TO TEBUCONAZOLE



# DATA ANALYSIS

The fungicide concentrations and corresponding mycelial growth inhibitions were used to calculate EC<sub>50</sub> using non-linear regression (Effective concentration inhibiting fungal growth by half)

	Fluxapyroxad	Pyraclostrobin	Tebuconazole
<i>Diaporthe gulyae</i> Shapiro-Wilk test	$p < 0.0001$	$p < 0.0001$	$p < 0.0001$
<i>Diaporthe helianthi</i> Levene's test	$p > 0.881$	$p > 0.859$	$p > 0.726$
Shapiro-Wilk test	$p < 0.0001$	$p < 0.0001$	$p < 0.0001$
Levene's test	$p > 0.713$	$p > 0.877$	$p > 0.822$



# EFFECTIVE CONCENTRATION INHIBITING FUNGAL GROWTH BY HALF

$$Y = E_0 + \frac{(E_{max} - E_0)}{1 + \left(\frac{\text{concentration}}{EC_{50}}\right)^{\text{Hill's coefficient}}}$$

- Y = expected response at a given fungicide concentration
- $E_{max}$  and  $E_0$  are the responses at maximum and zero fungicide concentration, respectively
- $EC_{50}$  is halfway between maximum and minimum response
- Hill's coefficient is the slope of the curve

# DATA ANALYSIS

*Diaporthe gulyae*

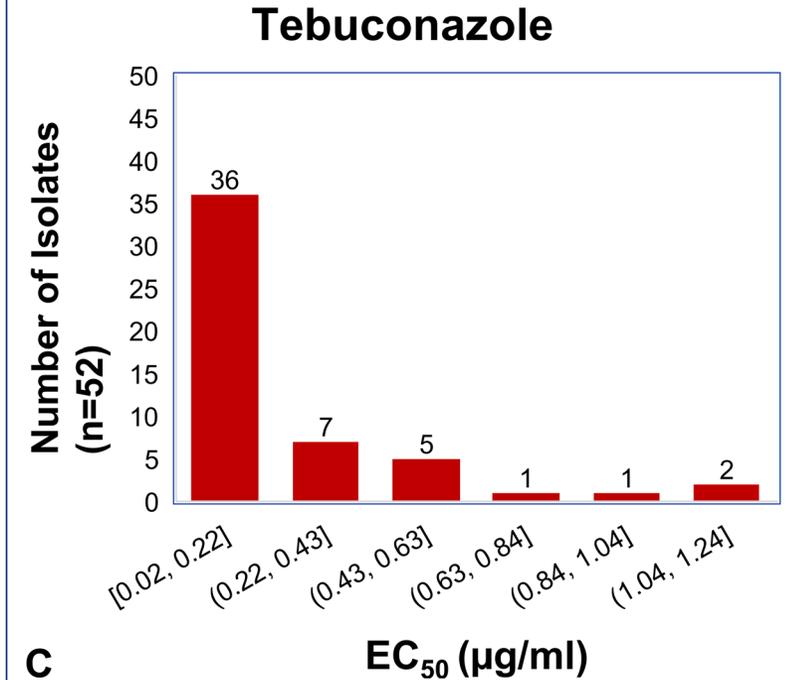
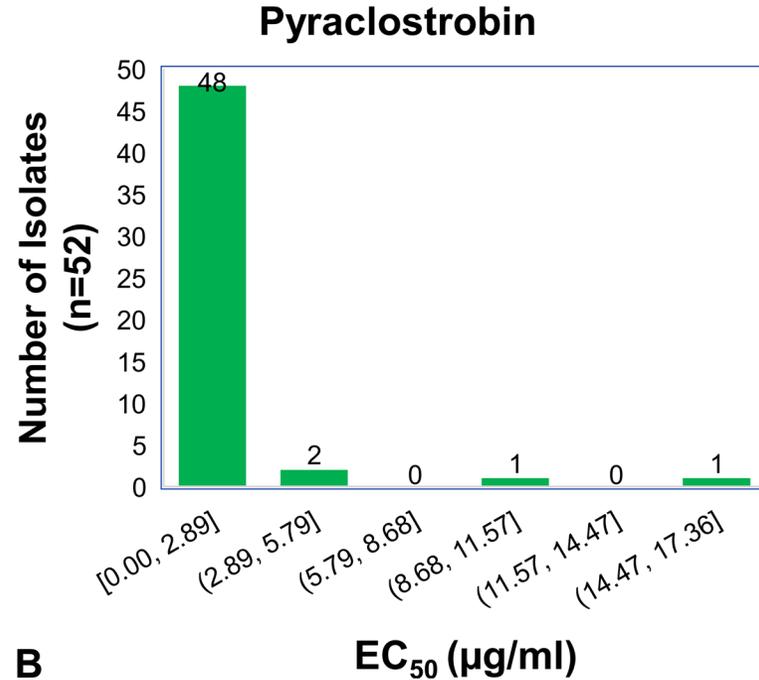
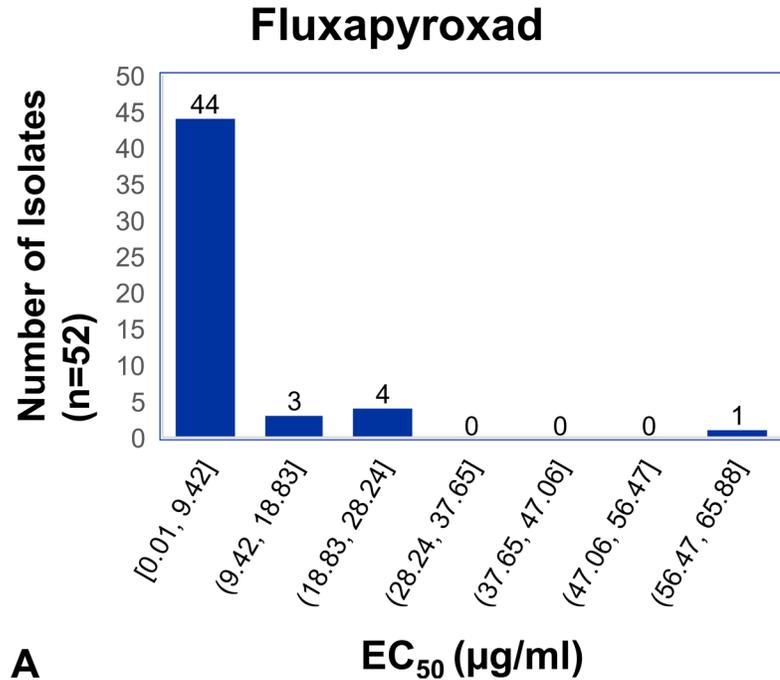
*Diaporthe helianthi*

	ATS value	df	p value
Fluxapyroxad	24.457	5.186	$p < 0.0001$
	44.985	5.170	$p < 0.0001$
Pyraclostrobin	11.588	5.066	$p < 0.0001$
	6.422	5.117	$p < 0.0001$
Tebuconazole	36.540	5.492	$p < 0.0001$
	14.635	5.356	$p < 0.0001$

Six, 22, and 21 isolates of *D. gulyae*, while three, three, and 13 isolates of *D. helianthi* had significantly greater EC50 ( $p < 0.0001$ ) than of the baseline isolate for fluxapyroxad, pyraclostrobin, and tebuconazole, respectively



Frequency distribution of effective fluxapyroxad (A), pyraclostrobin (B), and tebuconazole (C) concentrations that inhibited mycelial growth by 50% (EC<sub>50</sub>) for 52 isolates of *Diaporthe gulyae*.

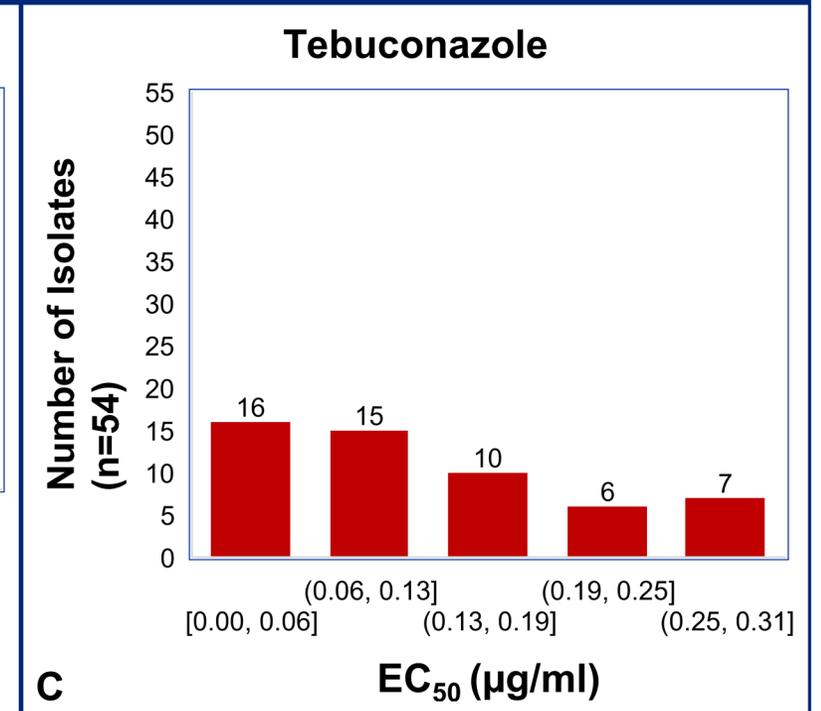
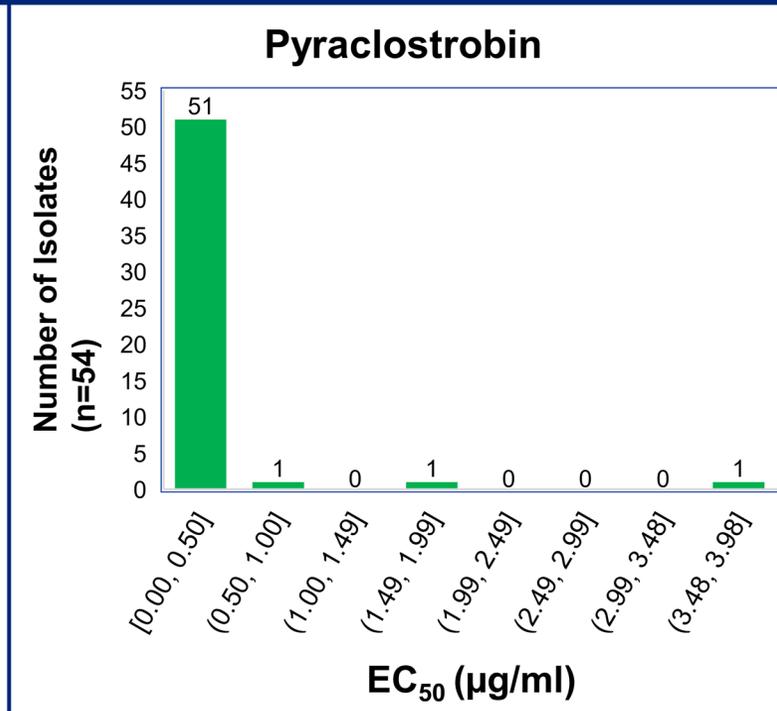
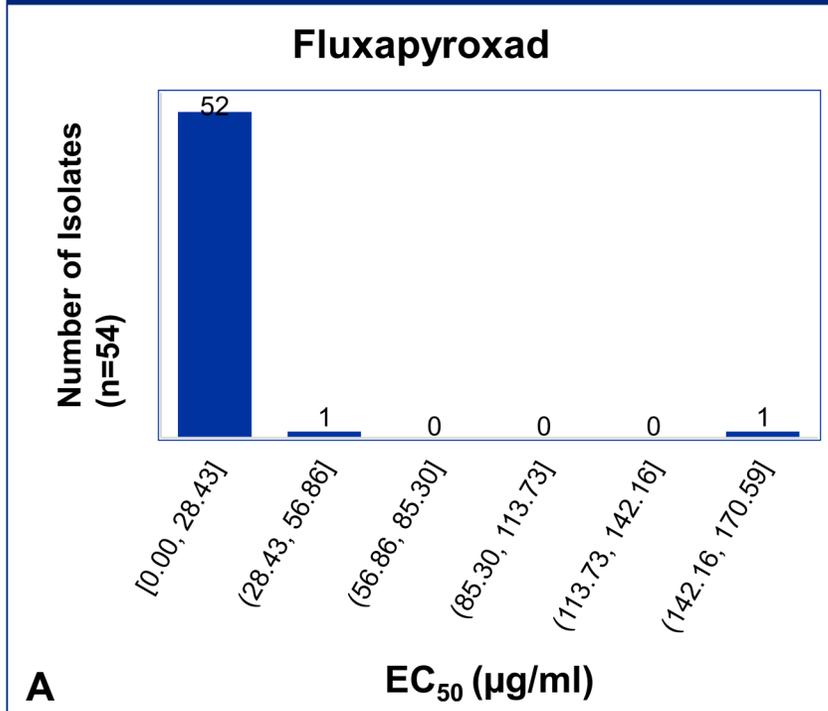


Mean EC<sub>50</sub> values were 6.234 (0.012 to 56.521) µg/ml for fluxapyroxad, 0.919 (0.001 to 17.358) µg/ml for pyraclostrobin, and 0.245 (0.0184 to 1.244) µg/ml for tebuconazole



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Frequency distribution of effective fluxapyroxad (A), pyraclostrobin (B), and tebuconazole (C) concentrations that inhibited mycelial growth by 50% (EC<sub>50</sub>) for 54 isolates of *Diaporthe helianthi*.



Mean EC<sub>50</sub> values were 5.999 (0.001 to 170.590) µg/ml for fluxapyroxad, 0.171 (0.001 to 3.980) µg/ml for pyraclostrobin, and 0.127 (0.002 to 0.313) µg/ml for tebuconazole



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# CORRELATIONS

Fungicide	Correlation coefficient	<i>p</i> value
Fluxapyroxad	0.874	$p < 0.0001$
Pyraclostrobin	0.984	$p < 0.0001$
Tebuconazole	0.880	$p < 0.0001$

There was a significant positive correlation between the EC<sub>50</sub> values of *D. gulyae* and *D. helianthi* for the three fungicides, indicating the fungicides have similar effect on the two fungi



# RESULTS

Possible decline in the sensitivity of *Phomopsis* species to fluxapyroxad, pyraclostrobin, and tebuconazole fungicides

*D. gulyae* and *D. helianthi* isolates exhibited a broad range of EC<sub>50</sub> values similar to other fungal pathogens:

*Botrytis cinerea* (0.07 to 7.1 µg/ml) for fluxapyroxad (Amiri et al. 2014) in strawberry

*Phomopsis asparagi* (0.009 to 0.153 µg/ml) for pyraclostrobin (Shi et al. 2020) in asparagus

*Fusarium graminearum* (0.0301 to 1.733 µg/ml) for tebuconazole (Anderson et al. 2020) in wheat

# IMPLICATIONS

The current study is the first multistate screening of *D. gulyae* and *D. helianthi* isolates for sensitivity to fluxapyroxad, pyraclostrobin, and tebuconazole in the United States

We established a protocol to monitor sensitivity of *D. gulyae* and *D. helianthi* to fungicides in future

Monitoring fungicide sensitivity is important to ensure prolonged and proper use of fungicides



# FUTURE WORK

Greenhouse  
testing

Cross sensitivity  
assays

Molecular assays  
for detection of  
mutations



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