

# **First report of *Diaporthe gulyae* systemic seed infection\* of sunflower – implications for DSC management**

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**\*paper submitted for review**



# The Australian *D. gulyae* outbreak triggered a seed infection investigation.....

- **2009/10** mid-stem lodging and yield losses
- Symptoms on leaves and stems almost identical to those of *D. helianthi* (*exotic*)
- Seed infection without visual symptoms on the capitulum
- Pycnidia emerged from the seed after incubation



Lodging and lesions typical of Phomopsis Stem Canker, seed infection in heavily infected plants

## ***Diaporthe gulyae*.....most virulent new species**

**Etymology:** In recognition of Dr. Tom Gulya (USDA-ARS) for his outstanding contributions to sunflower pathology research and enduring mentoring roles in the USA, Europe and Australia.



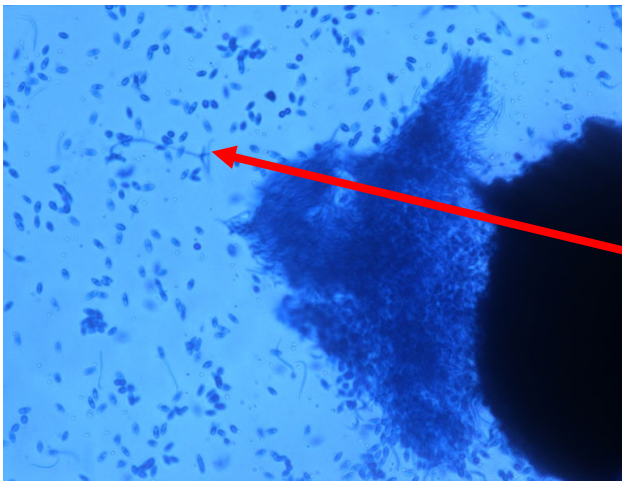
**First identified as a new species in Australia after a damaging stem canker outbreak on sunflower in 2009/10.**

**\*Described in 2011 (Thompson et al. 2011) as:  
*Diaporthe gulyae* RG Shivas, SM Thompson & AJ Young**

**\*Thompson SM, Tan YP, Young AJ, Neate SM, Aitken EAB, Shivas RG (2011a). Stem cankers on sunflower (*Helianthus annuus*) in Australia reveal a complex of pathogenic *Diaporthe* (*Phomopsis*) species. *Persoonia* 27: 80–89.**



**With the exception of conidia, *D. gulyae* (alpha conidia) symptoms on sunflower very similar to those of *D. helianthi* (beta conidia)....**



**Pycnidia in leaf, alpha conidia, *D. gulyae* type generated from leaf infection**



**Stem lesion colour gradient**



**Seed infection – oozing pycnidia**



# ***Stem lesions – the most commonly seen symptoms.***

***D. gulyae* stem symptoms similar to those of *D. helianthi*.  
Accurate differentiation of the species in the field is not recommended; Use molecular technologies to ID.**



## ***D. gulyae* virulence\* .....**

**\* Associated with 14 hosts in Australia – 5 crops and 9 weeds**

**\* Pathogenic on multiple crops: eg. safflower, sunflower, soybean, mungbean, lupin and \*\*peanut (papers in prep).**

**\* Pathogenic on multiple weeds: eg. *Xanthium* sp., *Physalis* sp., *H. annuus* (wild-type), *Sonchus oleraceus* (papers in prep).**

**Highly virulent on sunflower and other crops and weeds**

**Virulence rating 4 or 5 on a 1-5 rating scale**

***D. gulyae* isolates collected from leaves, stems, stem base/roots, capitula, seeds**

**\* PhD thesis: Brumpton Thompson S. 2020.**

*Diaporthe* species association with sunflower and other crops and weeds in eastern Australia. PhD Thesis, School of Agriculture and Food Services, The University of Queensland, Australia.

<https://doi.org/10.14264/uql.2020.779>

**\*\* Thompson et al. 2018**

**Stem slit inoculations – a harsh test, so consider the results with the biology of the *Diaporthe* genus in mind ..... opportunistic colonisation**

**Virulence ratings = degree of severity of infection**

Rating			Pathogenicity, virulence
1			Not pathogenic
2			Not pathogenic, or pathogenic with low virulence, or latent
3			Pathogenic, moderate virulence
4			Pathogenic, high virulence
5			Pathogenic, very high virulence

^pathogenic VR2-5, keep in mind plant age, time after inoc



***Diaporthe gulyae* isolates from a range of hosts – pathogenicity testing, lesion colour variation...**





# ***Diaporthe* spp (7) identified from sunflower seed in Australia.....virulence on sunflower (1-5 rating<sup>^</sup>)**

- ***D. ambigua*..... VR1/2**
- ***D. goulteri*..... VR2**
- ***D. gulyae*..... VR4/5**
- ***D. masirevicii*..... VR3**
- ***D. ueckeri* syn. *miriciae*..... VR3**
- ***D. serafiniae*..... VR2/3**
- ***Diaporthe* sp. nov. 14 (not tested)**



***D. gulyae***

<sup>^</sup>pathogenic VR2-5, keep in mind plant age, time after inoc

## ***Seed infection experiments.....***

- **1. Laboratory experiments – visual inspection of germination a range of *D. gulyae* infected seeds**
- **2. Glasshouse experiments – no other infected plants in G/H or within 20km**

**Match-stick inoculation of *D. gulyae* isolate at the 4<sup>th</sup> node**

**Stem lesions developed within 2 -5 dai**

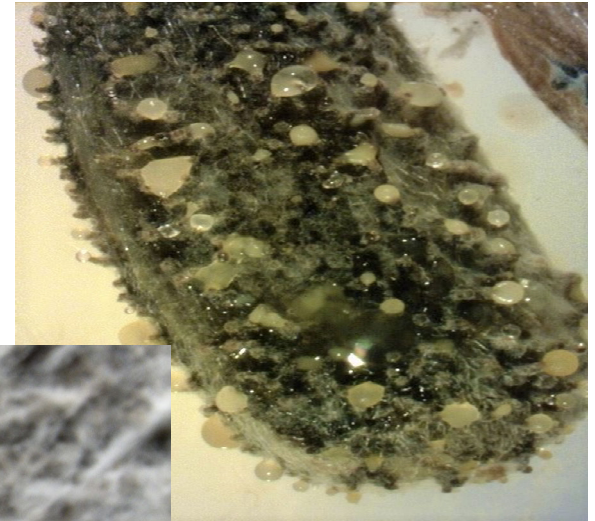
**Heads were bagged at budding**

**Plants harvested 21 days after physiological maturity and any head infection evaluated**

- **3. Glasshouse pot trials - evaluating emergence of infected seeds**

# 1. Lab assessment of germination of infected seeds.....

- **Highly infected seeds do not germinate on WAS**

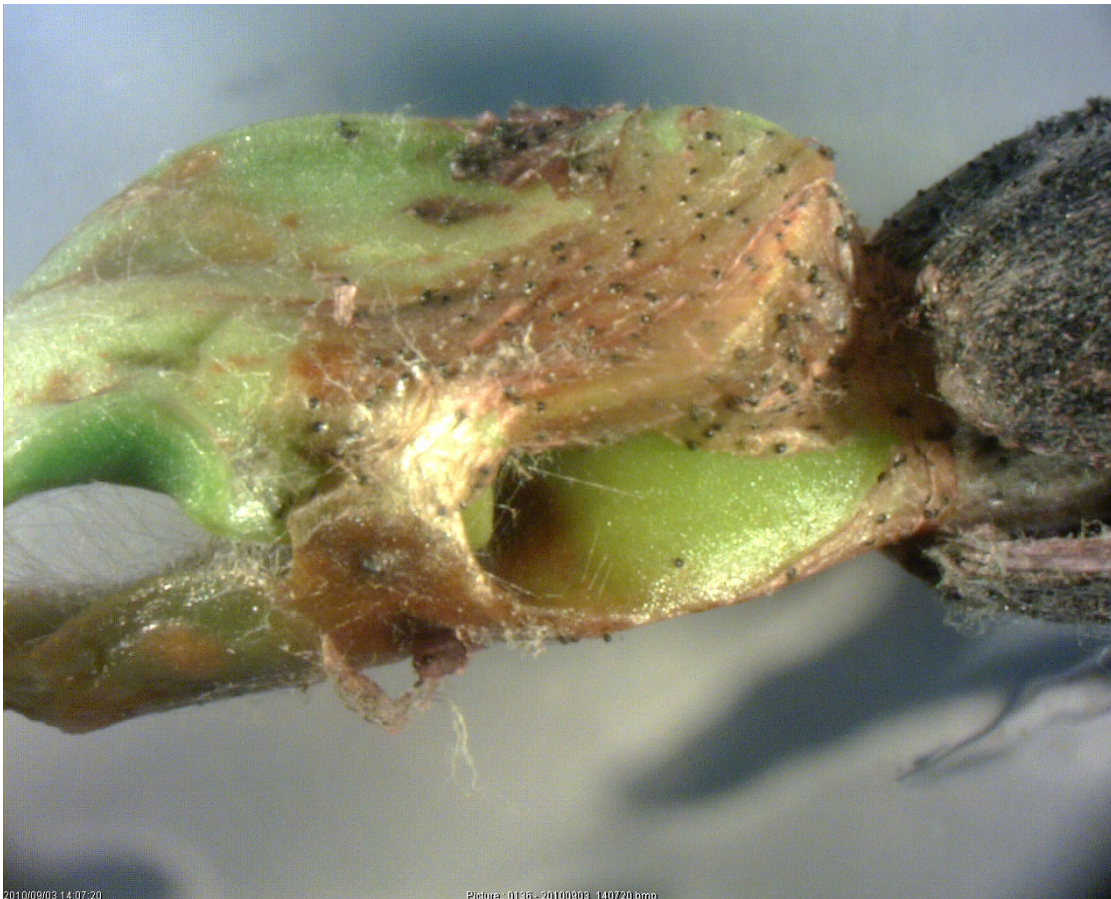




# 1. Infected seed germination (lab).....

**Some seeds (with fewer pycnidia observed) germinated on WAS then developed necrosis of cotyledons**

**Necrosis spread from the cotyledons to the emerging radicle**



## 2. Investigating systemic infection.....

- Sunflower plants grown in pots
- Inoculated at V10-R1 growth stage, match stick
- *D. gulyae* isolate of the type (BRIP 54025, MB561569)
- Progression of hyphal growth upwards in the stem was monitored by destructive sampling of a selection of inoculated plants
- Heads bagged during budding
- Surviving plants were harvested 21 days after physiological maturity
- Capitula pieces, seed and husks incubated



## **Proof of systemic infection via the stem.....**

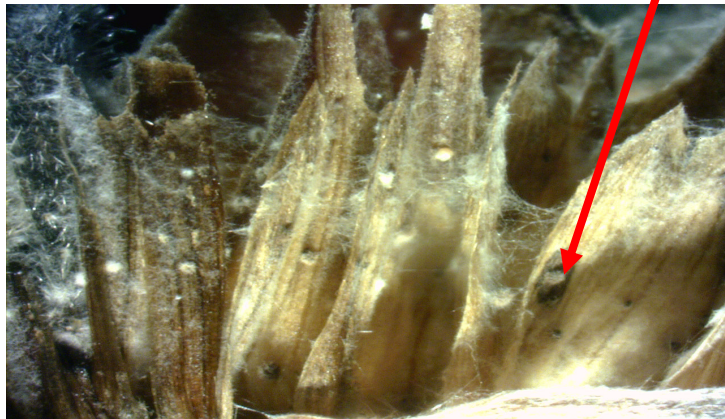
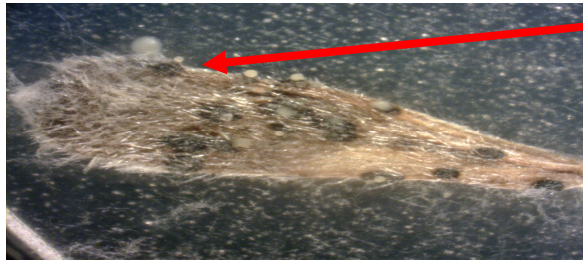
- **A lesion developed at the site of inoculation**
- **Lesions elongated over time**
- **Destructive sampling of the stems revealed hyphal colonisation of the stem tissues above the lesion**
- **Sampling occurred at +35, +49 and +56dai**
- **No signs or symptoms had developed on the capitula by the time of harvest at +21 days after phys maturity**
- **Harvested seeds were asymptomatic prior to incubation**



## **Results\* .....Capitula, husk and seed infection following stem inoculation (\*Paper submitted for review)**

- ***D. gulyae* hyphae and pycnidia emerged from incubated capitula pieces (approx. 71%), husks and seed (approx. 64%).**
- **Infected seeds were found in all surviving inoculated plants**

**No infection was seen in mock inoculated controls**



### 3. Infected seed germination and emergence in pot trials.....

- Most seeds with few pycnidia germinated and appeared disease free until V2-V3 in pot trials
- Some planted seeds emerged with infected pericarp intact and pycnidia viable
- Patches of brown necrotic tissue could be seen on the cotyledons of some emerged seedlings – *D. gulyae* isolated
- Such infected seeds are a possible inoculum reservoir and source of new outbreaks
- Further investigation of *D. gulyae* colonisation of emerged seedlings and older plants suggested



# Sunflower seedling emergence habit may assist *D. gulyae* survival on emerging seedlings...

- **Epigeal emergence - cotyledons are pulled rather than pushed (hypogeal) from the soil**
- **Then cotyledons are extended above the soil surface by elongation of the hypocotyl**
- **Pericarp halves (whether infected or not) can be retained on emerged cotyledons or dropped onto the soil surface after emergence.**





## **Seed infection of *D. gulyae* vs *D. helianthi***

- **Both species are known to infect sunflower seeds**
- **Unclear whether *D. helianthi* infects capitulum directly (ascospores) - some reports of this occurring.**
- **Also reports suggesting systemic seed infection of *D. helianthi* via the stem**
- **Opinions vary as to the significance of *D. helianthi* seed infection in broadening distribution**
- **Our results are clear – *D. gulyae* can infect seeds systemically following stem infection**
- **Some infected seeds can germinate and produce a viable seedling**
- **Infection on emerged cotyledons can occur**
- **Viable *D. gulyae* pycnidia were found on attached pericarp pieces after seedling emergence**
- ***D. gulyae* seed infection has the potential to broaden distribution**

## ***Implications for management.....***

**Growers and advisors, seed companies – know the biology of pathogens, symptoms and timing of infection; understand the survival strategies of each pathogen; be vigilant, know your crops and paddocks, be aware of the disease levels and biology of your pathogens**

**Have a Whole of Farming System Overview**

**Seed companies – vigilance, inspect nurseries, bury or process infected stubble, be aware of any disease outbreaks; utilize seed testing**

**Harvest sunflower as soon as possible after physiological maturity**

**Seed infection has the potential to broaden distribution of *D. gulyae* to new fields, regions, countries**

**Importance of biosecurity protocols**

# Suggested further studies.....

- ***D. gulyae* ascospore production and infection of the capitulum needs further investigation**
- **Investigate possible *D. gulyae* endophytic colonisation of seedlings emerged from infected seeds**
- **Evaluate temperature, humidity and timing of infection parameters vs incidence and/or severity of seed infection (similar to those identified for soybean?)**
- **Investigate *D. gulyae* hyphal colonisation of host tissues – same as for *D. helianthi*?**
- **Investigate *D. gulyae* hyphal and pycnidial development within the seed.**
- **Numbers of pycnidia vs seed germination**
- **Fungicides/ seed treatments**





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## **Publications.....**

**Thompson SM, Tan YP, Young AJ, Neate SM, Aitken EAB, Shivas RG (2011a). Stem cankers on sunflower (*Helianthus annuus*) in Australia reveal a complex of pathogenic *Diaporthe (Phomopsis)* species. *Persoonia* 27: 80–89.**

**Thompson SM, Tan YP, Shivas RG, Neate SM, Morin L, Bissett A, Aitken EAB (2015). Green and brown bridges between weeds and crops reveal novel *Diaporthe* species in Australia. *Persoonia* 35: 39-49.**

**Thompson S, Grams R, Neate S, Shivas R, Ryley M, Tan YP, et al. (2018a). First reports of *Diaporthe kongii*, *D. masirevicii* and *D. ueckerae* associated with stem and peg dieback on peanut in Australia. *Plant Disease*: 102: 1459.**

**Thompson SM, Tan YP, Neate SM, Shivas RG, Lindbeck K, Aitken EAB (2018b). *Diaporthe novem* isolated from sunflower (*Helianthus annuus*) and other crop and weed hosts in Australia. *European Journal of Plant Pathology* 152: 823-831.**

**Brumpton Thompson S. 2020. *Diaporthe* species association with sunflower and other crops and weeds in eastern Australia. PhD Thesis, School of Agriculture and Food Services, The University of Queensland, Australia. <https://doi.org/10.14264/uql.2020.779>**