

2016 Progress on Transferring Sclerotinia Resistance Genes from Wild *Helianthus* Species into Cultivated Sunflower

Zhao Liu (NDSU)

PI: Dr. Chao-Chien Jan (USDA-ARS, NCSL)

Co-PI: Gerald J. Seiler (USDA-ARS, NCSL)

Cooperators: Khalid Y. Rashid (Agric. & Agri-Food Canada)

Xiwen Cai (NDSU)

INTRODUCTION

- Sclerotinia, also called white mold, is caused by *Sclerotinia sclerotiorum* (Lib.) de Bary
- It is a severe and widespread fungal disease attacking 64 plant families, and more than 360 species, causing both yield and quality reduction
- Moreover, species of the genus *Sclerotinia* can function either as soilborne or airborne pathogens
- It is difficult to control because it is a long-lived pathogen that is not host-specific, and the resistance is controlled by multiple genes
- Wild *Helianthus* species are important genetic resources for the improvement of cultivated sunflower

53

Helianthus Species

14 Diploid annuals ($2n=2x=34$)

26 Diploid perennials ($2n=2x=34$)

3 Tetraploid perennials ($2n=4x=68$)

7 Hexaploid perennials ($2n=6x=102$)

1 Mixploid perennials ($2n=2x=34, 4x=68$)

2 Mixploid perennials ($2n=4x=68, 6x=102$)

Wild *Helianthus* species resistant to sunflower diseases

Diseases	Wild species sources
Rust	<i>H. annuus</i> ; <i>H. petiolaris</i> ; <i>H. argophyllus</i>
Downy mildew	<i>H. annuus</i> ; <i>H. petiolaris</i> ; <i>H. praecox</i> ; <i>H. hirsutus</i> ; <i>H. laevigatus</i> ; <i>H. californicus</i> ; <i>H. ciliaris</i> ; <i>H. pauciflorus</i> ; <i>H. resinosus</i> ; <i>H. strumosus</i> ; <i>H. tuberosus</i> ; <i>H. x laetiflorus</i> ; <i>H. smithii</i>
<u>Sclerotinia</u>	<i>H. agrestis</i> ; <i>H. argophyllus</i> ; <i>H. debilis</i> ; <i>H. neglectus</i> ; <i>H. petiolaris</i> ; <i>H. praecox</i> ; <i>H. californicus</i> ; <i>H. ciliaris</i> ; <i>H. eggertii</i> ; <i>H. pauciflorus</i> ; <i>H. resinosus</i> ; <i>H. tuberosus</i> ; <i>H. hirsutus</i> ; <i>H. maximiliani</i> ; <i>H. nuttallii</i> ; <i>H. giganteus</i> ; <i>H. grosseserratus</i> ; <i>H. salicifolius</i> ; <i>H. smithii</i>
Phomopsis brown stem canker	<i>H. debilis</i> ; <i>H. argophyllus</i> ; <i>H. pauciflorus</i> ; <i>H. tuberosus</i> ; <i>H. hirsutus</i> ; <i>H. pauciflorus</i> ; <i>H. maximiliani</i> ; <i>H. nuttallii</i> ; <i>H. mollis</i> ; <i>H. occidentalis</i> ; <i>H. divaricatus</i> ; <i>H. resinosus</i> ; <i>H. strumosus</i>
Alternaria leaf spot	<i>H. praecox</i> ; <i>H. debilis</i> subsp. <i>cucumerifolius</i> ; <i>H. debilis</i> subsp. <i>silvestris</i> <i>H. tuberosus</i> ; <i>H. x laetiflorus</i>
Powdery mildew	<i>H. debilis</i> subsp. <i>silvestris</i> ; <i>H. praecox</i> subsp. <i>praecox</i> ; <i>H. bolanderi</i> ; 14 perennials
Rhizopus head rot	<i>H. divaricatus</i> ; <i>H. hirsutus</i> ; <i>H. x laetiflorus</i> ; <i>H. resinosus</i>
Phoma black spot	<i>H. argophyllus</i> ; <i>H. maximiliani</i> ; <i>H. tuberosus</i> ; <i>H. pauciflorus</i>
Charcoal rot	<i>H. tuberosus</i> ; <i>H. mollis</i> ; <i>H. maximiliani</i> ; <i>H. resinosus</i> ; <i>H. pauciflorus</i>
Broomrape	<i>H. anomalus</i> ; <i>H. exilis</i> ; all perennial species

Note: Wild annuals are in black, and wild perennials are in red.

Sources: Block CC et al. American Phytopathological Society Abstracts 102:S4.12 (2012)

Christov M. Proc. 17th International Sunflower Conference, pp 709-714 (2008)

Christov M. et al. *Helia*, 32: 65-74 (2009)

Seiler G. Book chapter: Utilization of wild *Helianthus* species in sunflower breeding, pp 355-429 in Kovacevic Z (Ed.) Sunflower Genetics and Breeding: International Monography.(2012)

OBJECTIVES

- ❑ Identify resistant wild perennial *Helianthus* species populations and interspecific amphiploids
- ❑ Transfer resistance genes into a cultivated background
- ❑ Study the inheritance of resistance
- ❑ Release *Sclerotinia* head and stalk rot resistant germplasm

MATERIALS

□ Phase 1 (since 2004)

- ❖ 2 hexaploids (backcrossed with HA 410)
H. californicus and *H. schweinitzii*
- ❖ 3 diploids (backcrossed with HA 410)
H. maximiliani, *H. giganteus*, and *H. grosseserratus*
- ❖ 2 diploids (backcrossed with HA 441)
H. nuttallii and *H. maximiliani*
- ❖ 5 amphiploids (backcrossed with HA 410)
H. nuttallii/P21, *H. maximiliani*/P21, *H. gracilentus*/P21,
H. grosseserratus/P21, and *H. strumosus*/P21

□ Phase 2 (since 2011)

- ❖ 1 hexaploid (backcrossed with HA 410)
H. resinusus
- ❖ 1 tetraploid (backcrossed with HA 410, HA451, NMS HA 89)
H. hirsutus
- ❖ 4 diploids (backcrossed with HA 410, HA451, NMS HA 89)
H. salicifolius, *H. occidentalis* subsp. *plantagineus*, *H. silphioides*, and
H. divaricatus

□ Phase 3 (since 2013)

- ❖ 2 hexaploids (backcrossed with HA 410, NMS HA 89)
H. strumosus and *H. tuberosus*
- ❖ 1 tetraploid (backcrossed with HA 410, NMS HA 89)
H. decapetalus
- ❖ 1 diploid (backcrossed with HA 410, NMS HA 89)
H. simulans

□ Phase 4 (added in 2016)

- ❖ 1 amphiploid (backcrossed with HA 410, NMS HA 89)
H. atrorubens/HA 89
- ❖ 3 hexaploids (backcrossed with HA 410, NMS HA 89)
H. smithii, *H. laevigatus*, and *H. pauciflorus* (*rigidus*)

Note: (1) For most species, 2-3 accessions were used.
(2) The materials in red were continued in 2016.

METHODS

- Traditional crossing and backcrossing
- Embryo rescue
- Mitotic chromosome counting
- Pollen fertility examination
- Seed increase in field and greenhouse
- Head and stalk rot field test
- Stalk rot greenhouse test
- GISH and FISH
- Molecular markers: SSR, SNPs

RESULTS AND DISCUSSION

Table 1. Seed increases for progeny families in 2012-2016

Sources	2012	2013	2014	2015	2016
Hexaploid	32	36	52	39	20
Diploids-HA 410	55	53	6	110	38
Diploids-HA 441	120	36	96	72	24
Amphiploids	34	42	17	157	16
New diploids*		370	19	108	21
New tetraploids*		60	4	66	1
New hexaploids*					32
Total	241	597	194	552	152

- * Phase 2 and 3 crosses.
- Seed increase started in 2008.



Table 2. Field test of progeny families in 2012-2016

Sources	Stalk rot (SR)					Head rot (HR)				
	2012	2013	2014	2015	2016	2012	2013	2014	2015	2016
Hexaploid	65	43	39	9	9	27	1	1	0	0
Diploids-HA 410	56	27	25	18	18	45	11	11	6	3
Diploids-HA 441	121	51	51	5	5	90	41	41	14	7
Amphiploids	42	18	16	7	7	11	0	0	0	0
New diploids			368	318	30			110	316	35
New tetraploids			60	54	6			0	0	0
Total	284	139	559	411	75	173	53	163	336	45

- Disease evaluations started in 2009.



Table 3. Third year test of materials from Phase I crosses for head rot in 2016

Pedigree*	Carrington, ND 2016		Staples, MN 2016	
	Disease Severity	Disease Incidence	Disease Severity	Disease Incidence
TEST 2 (Second Retest)	0-5	%	0-5	%
((NMS HA 89 x GRO PI613793) HA 410*2), BC2F3 (47)	3.50	83	1.76	42
((NMS HA 89 x GRO PI613793) HA 410), BC2F3 (66) →	1.83	38	1.94	44
((NMS HA 89 x GRO PI613793) HA 410), BC1F4 *new 69*	2.89	65	2.96	69
→ Recurrent parent HA 410	4.88	92	3.71	89
((NMS HA 89 x 1323(MAX) x HA 441), BC1F5 (7) →	0.94	22	0.92	25
((NMS HA 89 x 1324(NUT) x HA 441), BC1F5 (8) →	0.84	24	1.36	34
((NMS HA 89 x 1008 (NUT)) x HA 441) HA 441, BC2F4 (9)	2.54	59	1.32	33
((NMS HA 89 x 1018 (MAX)) x HA 441), BC1F6 (10) →	1.34	37	0.93	20
((NMS HA 89 x 1324 (NUT) x HA 441), BC1F5 (11) →	1.13	40	0.37	11
((NMS HA 89 x 1008 (NUT)) x HA 441) HA 441), BC2F4 (15) →	1.60	47	2.13	52
((NMS HA 89 x 1008 (NUT)) x HA 441) HA 441), BC2F4 (16)	2.58	55	1.82	54
→ Recurrent parent HA 441	1.39	42	2.15	57
Checks				
Susceptible check HA 89 (S)	3.86	83	3.26	82
Susceptible check Cargill/Mycogen (270/272) (S)	2.13	54	4.04	82
Resistant check Croplan 305 (R)	1.86	41	1.95	47
Resistant check Croplan 343 (R)	0.87	3	0.95	27

→ Entries close to or better than resistant check

→ Entries close to or better than recurrent parent

Table 4. Second year test of materials from Phase 2 crosses for head rot in 2016







Pedigree*	Carrington, ND 2016		Staples, MN 2016	
	Disease Severity	Disease Incidence	Disease Severity	Disease Incidence
TEST 3 (First Retest)	0-5	%	0-5	%
NMS HA 89 x (SAL x HA 410), F2 (1011) 	1.94	52	1.18	30
NMS HA 89 x (SAL x HA 410), F2 (1014) 	2.20	51	1.61	43
NMS HA 89 x (SAL x HA 410), F2 (1063)	4.17	85	3.08	77
NMS HA 89 x (SAL x HA 410), F2 (1069)	3.89	84	2.26	58
NMS HA 89 x (SAL x HA 410), F2 (1072) 	1.69	44	1.40	36
NMS HA 89 x (SAL x HA 410), F2 (1074) 	2.23	51	2.50	53
NMS HA 89 x (SAL x HA 410), F2 (1077)	3.94	82	3.67	89
NMS HA 89 x (OCC x HA 410), F2 (1047) 	1.74	40	1.91	43
NMS HA 89 X (OCC X HA 410), F2 (1085)	3.68	84	1.61	41
NMS HA 89 X (OCC X HA 410), F2 (1097)	3.50	73	2.60	56
NMS HA 89 X (OCC X HA 410), F2 (1098)	4.59	95	3.24	79
 Recurrent parent HA 410	4.41	90	2.68	68
Checks				
Susceptible check HA 89 (S)	3.86	83	3.26	82
Susceptible check Cargill/Mycogen (270/272) (S)	2.13	54	4.04	82
Resistant check Croplan 305 (R)	1.86	41	1.95	47
Resistant check Croplan 343 (R)	0.87	3	0.95	27

Table 5. First year test of materials from Phase 2 crosses for head rot in 2016

Pedigree*	Carrington, ND 2016		Staples, MN 2016	
	Disease Severity	Disease Incidence	Disease Severity	Disease Incidence
TEST 4 (New Selections)				
NMS HA 89 x (SAL x HA 410), F2 (1120) →	2.59	47	1.90	44
NMS HA 89 x (SAL x HA 410), F2 (1121) →	2.00	52	0.69	21
NMS HA 89 x (SAL x HA 410), F2 (1137)	4.05	86	2.97	71
NMS HA 89 x (SAL x HA 410), F2 (1138)	4.12	98	2.13	68
NMS HA 89 x (SAL x HA 410), F2 (1139)	4.25	85	2.19	54
NMS HA 89 x (OCC x HA 410), F2 (1163) →	2.39	54	2.18	48
NMS HA 89 x (OCC x HA 410), F2 (1195)	3.53	75	2.04	51
NMS HA 89 x (OCC x HA 410), F2 (1208) →	1.92	58	0.68	51
→ Recurrent parent HA 410	4.24	85	2.73	60
Checks				
Susceptible check HA 89 (S)	3.86	83	3.26	82
Susceptible check Cargill/Mycogen (270/272) (S)	2.13	54	4.04	82
Resistant check Croplan 305 (R)	1.86	41	1.95	47
Resistant check Croplan 343 (R)	0.87	3	0.95	27

*The first three letters of the *Helianthus* species are used to identify the species source: GRO=*H. grosseserratus*; MAX=*H. maximiliani*; NUT=*H. nuttallii*; SAL=*H. salicifolius*; and OCC=*H. occidentalis*.

(S)= Susceptible; (R)=Resistant. The number in parentheses at the end of each pedigree is the family ID of the selected family.

Table 6. Backcrosses between wild perennials and cultivated sunflower-Phase 3

Parentage	BC ₁ F ₁ (2014 and 2016)				BC ₂ F ₁ (2015)			
	Plants	Fertility %	BC ₂ F ₁ seeds	Seed set %	Plants	Fertility %	BC ₃ F ₁ /BC ₂ F ₂ seeds	Seed set %
<i>H. strumosus</i> (PI 547217) × HA 410	29	3.01	24 ^a 3 ^b	0.13 0.05	7	56.01	290	9.67
<i>H. strumosus</i> (PI 547226) × HA 410	25	4.48	9 ^a 0 ^b	0.12 0.00	3	77.49	43	2.90
<i>H. tuberosus</i> (PI 547242) × HA 410	22	7.48	11 ^a 8 ^b	0.14 0.15	4	7.80	34	1.77
<i>H. tuberosus</i> (PI 650089) × HA 410	21	3.07	64 ^a 14 ^b	0.91 0.24	32 ^a 9 ^b	- 33.56	Many 527	76.78 12.61
<i>H. tuberosus</i> (PI 650105) × HA 410	20	5.00	23 ^a 901 ^b	0.28 14.18	4 ^a 6 ^b	15.28 -	56 50	3.86 2.59

^a Obtained by crossing HA 410 pollen to BC₁F₁ plants.

^b Obtained by crossing BC₁F₁ pollen to NMS HA 89.

Backcrosses between wild perennials and cultivated sunflower-Phase 4

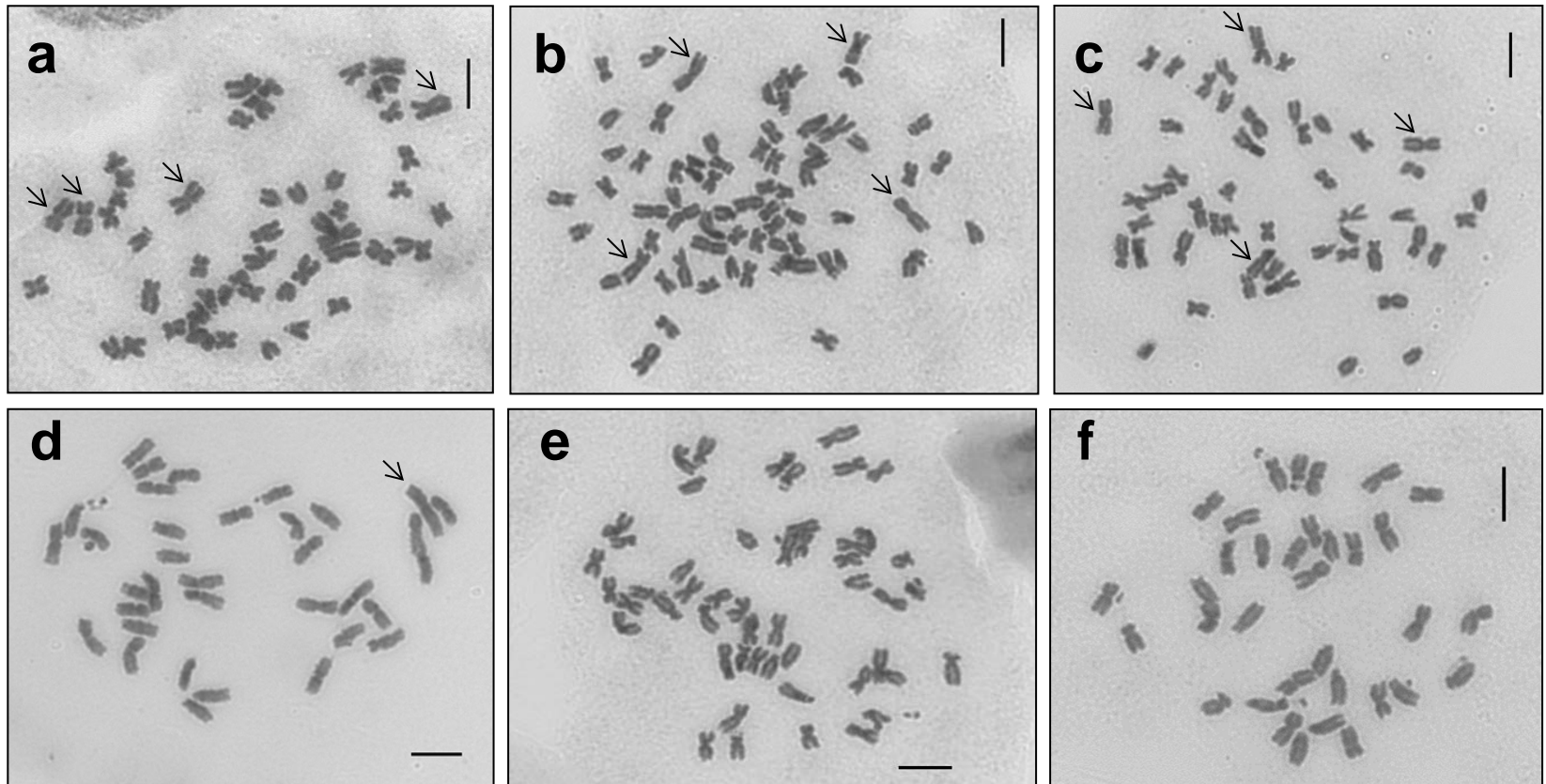


Figure 1. Chromosome spreads of several backcross progenies derived from *H. atrorubens* (a-d) and *H. laevigatus* (e and f). (a) G95/72 x HA 89, BC₁F₁, 2n=51; (b) G16/1039 x HA 410, BC₁F₂, 2n=68; (c) G16/1039 x HA 410, BC₂F₁, 2n=49; (d) NMS HA 89 x G16/1036-46, BC₂F₁, 2n=35; (e) G16/1053 x HA410, BC₂F₁, 2n=50; (f) NMS HA 89 x G16/1053-64, BC₂F₁, 2n=34. Notice that there are large chromosomes (arrows) from *H. atrorubens* in the cultivated background in Figures a-d. Bars=5 μ m.

Table 7. Sclerotinia resistance germplasms crossed with HA 234 for RIL population development

Germplasms	Source	Pedigree for female parent	Selfed/Sib seed set (%)	Crossed seed set (%)
HR-MAX	09/4008	(NMS HA 89 x 1018 (MAX)) x HA 441 ³ , BC3F3	43.3	55
HR-NUT	09/4041	(NMS HA 89 x 1008 (NUT)) x HA 441, BC1F5	65	56.7
BSR-MAX	09/4011	(NMS HA 89 x 1018 (MAX)) x HA 441 ³ , BC3F3	15	26.9
BSR-NUT	10/4144	(NMS HA 89 x 1008 (NUT)) x HA 441, BC1F4	55	67
BSR-DIV	11/4484	[(DIV(68) x GRO (68)) x HA 410 ³ , BC2F2] x HA 410, BC3F2	43	65
BSR-STR	11/4489	[STR (68) x HA 410 ³ , BC2F2] x HA 410, BC3F2	51	55
BSR-CAL	09/4271	CAL 2376 x HA 410 ⁵ , BC4F4	-	20

- (1) The germplasm release for these resistant sources are in progress.
- (2) Root tips, leaf samples and pollen were collected from these sources.
- (3) 1-3 plants from each resistant source were emasculated and crossed with cultivated HA 234 in 2016.

Table 8. Resistant and susceptible entries to stalk rot derived from *H. californicus* (CAL) for GISH and/or GBS analyses

Entry #	Source	Pedigree	SR Score (%)	Selfed seed set (%)
28	09/4104	CAL 2376 x HA 410 ⁵ , BC4F4	0	25.00
216	10/4386	CAL 2376 x HA 410 ⁵ , BC4F4	0	67.5
235	11/4470	CAL 2376 x HA 410 ⁵ , BC4F4	0	61.25
258	11/4478	CAL 2376 x HA 410 ⁵ , BC4F4	0	27.54
304	09/4306	CAL 2376 x HA 410 ⁵ , BC4F4	0	18.33
29	09/4106	CAL 2376 x HA 410 ⁵ , BC4F4	8-63	40.00
30	09/4110	CAL 2376 x HA 410 ⁵ , BC4F4	0-67	41.25
32	09/4114	CAL 2376 x HA 410 ⁵ , BC4F4	0-56	45.00
41	09/4163	CAL 2376 x HA 410 ⁵ , BC4F5	0-50	47.50
228	11/4466	CAL 2376 x HA 410 ⁵ , BC4F4	33-45	34.21
317	11/4497	CAL 2376 x HA 410 ⁵ , BC4F5	7-46	72.50

- (1) These entries were selected based on field screening in 2009-2015.
- (2) Root tips were collected for GISH analysis.
- (3) Leaf samples were collected for GBS analysis.

SUMMARY

- In the last 12 years, the project has utilized 21 wild perennials, produced ~3100 progeny families, tested ~2500 families for stalk rot, ~1400 families for head rot in the field (including retests)
- The germplasm release for seven *Sclerotinia* resistant bulks are in progress
- In 2016, field evaluation identified several families derived from six diploid perennials with good resistance to head rot
- Seed was increased in the field for more than 150 progeny families
- The BC₁F₁ progenies (2n=51) derived from four perennials were advanced to the BC₂F₁ generation
- RIL population development was initiated for *Sclerotinia* resistance QTL mapping

FUTURE WORK

- ✓ **Thirteen amphiploids** derived from eight wild perennial species will be released as germplasms
- ✓ **Additional families with better resistance** than the recurrent parents identified in the different trials will be retested in 2017
- ✓ Continue to backcross the progenies derived from different sources to **reduce $2n$ to 34**
- ✓ **Continue to develop QTL mapping populations by RIL and doubled-haploid approaches**
- ✓ GISH and GBS comparisons between resistant and susceptible progenies derived from *H. californicus*
- ✓ Identify chromosome **addition lines, and characterize alien chromosomes or fragments** in cultivated background utilizing GISH and FISH techniques and molecular markers

ACKNOWLEDGEMENTS

Funding: National Sclerotinia Initiative

Lisa Brown

Dr. Jiujuan Feng

Dr. Hongxia Wang

Dr. Jichong Zhang

Puying Zheng

Marjorie Olson

Megan Ramsett

Leonard Cook (Retired)

Angelia Hogness

Dr. Lili Qi

Dr. William Underwood

Dr. Thomas Gulya (Retired)

Dr. Nikolay Balbyshev (Retired)

Dr. Zahirul Talukder

Dr. Guojia Ma

Dr. Yunming Long

Chris Misar

Michelle Gilley

Dr. Charles Block (USDA-ARS, North Central Regional Plant Introduction Station, Ames, IA) (Retired)

Many others who have helped !